

## MYCOBIOTA ASSOCIATED WITH CAMEL HAIR AT TAIZ CITY, YEMEN

A.M. H. Sallam\*, Gamal A. AL-Ameri\*\*

Applied Microbiology Department, Faculty of Science\*, Microbiology Department, Faculty of Medicine\*\*, Taiz University, Taiz, Yemen

#### **ABSTRACT:**

The mycobiota of 50 camel hair samples collected from different localities in Taiz city were estimated using the soil plating technique and Sabouraud's dextrose agar at 28° C. Forty-three species belonging to 15 genera were collected. The most prevalent genera were: *Chysosporium, Aspergillus* and *Scopulariopsis*. Some dermatophytic species were also recovered and these were represented by *Microsporum canis*, M. *gypseum Trichophyton rubrum* and *T. verrucosum*. Camel hair seem to represent an adequate reservoir for several pathogenic fungi.

Kew words: Animal hairs, dermatophytes, keratinophilic fungi, camel.

### **INTRODUCTION:**

Animals are known to play an important role in the epidemiology of both animal and human mycoses. They can act as a reservoir or vectors for these diseases (Rippon, 1982; Ogbonna et al., 1986; and Ali- Shtayeh et al., 1989).

*Keratinophilic fungi* have received considerable attention in recent years. These organisms include *dermatophytes* which are able to degrade various types of keratinous substances. These substances occurr in nature mainly in the form of hairs, wools, feathers, horns, hooves, nails, skin and other cornified appendages and constitute natural baits for these *keratonophilic fungi* (Khanam and Jain, 2002; Singh et al., 2009).

*Keratinophilic fungi* include a variety of filamentous fungi belonging mainly to hyphomycetes and several other taxonomic groups. Hyphomycetes include *dermatophytes* and a great variety of non- dermatophytic filamentous fungi (Mukesh and Sharma, 2010). *Keratinophilic fungi* can be considered as potential pathogens (Marcella and Mercantini, 1986).

Several studies have been conducted on mycobiota associated with hair of different kinds of animals in many parts of the world (Moses and Sunday, 2001; Shukia, et al. 2003; Periasamy et al., 2004; Dobrowolska et al., 2006; Ulfig, 2007; Maghraby et al., 2008; Mandeel et al., 2009; Nichita and Marcu, 2010; Rostami, et al., 2010; Sallam and Alkolaibe, 2010; Shokri and Khosravi, 2011; and Kraemer et al., 2012).

In Yemen there are no records available on *dermatophytes* and *keratinopilic fungi* associated with camel hair. The aim of the present work was to survey those fungi and identify the mycobiota associated with camel hair which might be causal agents for human mycosis..

#### **MATERIALS AND METHODS:**

#### The areas of study:

Hair-baiting technique was used for isolation of keratinophilic fungi as employed by Ress (1967) with some modifications. Fifty samples of camel hair were collected from different localities at Taiz city, Yemen during the period from October 2011 to December 2012. These samples were placed in clean plastic bags and transferred immediately to the laboratory. Five hair segments were placed on dry sterile clay soil (25- 50 g) in each sterile Petri-dish (3 plates for each sample) moistened with sterilized distilled water (20-25% moisture content) and remoistened whenever necessary. The plates were incubated at 28°C for 10-12 weeks. The moulds which appeared on hair fragments were transferred to the surface of Sabouraud's dextrose agar medium (Moss and McQuown, 1969), supplemented with chloramphenicol (0.5 mg/ml) and

cyclohexamide (0.5 mg/ml). Cultures were then incubated at 28°C for 2-3 weeks and the developing fungi were counted, identified (based on morphological and microscopic characters) following the keys of Moubasher (1993) and de Hoog et al. (2000). Fungal colonies were calculated per 15 hair fragments for each sample.

#### **Results and Discussion :**

A total of 34 species appertaining to 15 genera were recovered from camel hair samples (Table 1). These fungi included *dermatophytes* such as *Trichophyton*, and *Microsporum*, as well as true *keratinophilic* fungal species belonging to *Chrysosporium* and *Scopulariopsis*. Other *cycloheximide*- resistant were also recorded.

Chrysosporium was the most frequent genus being recovered from 62% of samples and representing 41% of all fungal isolates. This genus was also isolated from goat and sheep hairs in Libya by El-Said et al. (2009) who found that Chrysosporium was recorded in 92% and 96% of the samples and 91.2% and 87.8% of the total fungi of goat and sheep hairs, respectively. In Egypt, Bagy and Abdel-(1985) indicated Hafez also that Chrysosporium was the most frequent genus of the camel (98.3%) and goat (91.7%) hair samples from Al-Arish. Also, Abdel-Gawad (1997) observed that Chrysosporium was the most common fungus on sheep wool. In Italy, Marcella et al. (1985) found that, out of 115 animals examined, 54 presented keratinophilic fungi of which Chrysosporium spp. were the

most common. In the current study Chrysosporium was represented by four species, of which C. tropicum was the most frequent. It was found in 46% of the samples, and comprised 23% of all isolates. The remaining Chrysosporium species (C. indicum, C. keratinophilum, and C. georgii) were isolated in low frequency of occurrence. They were identified from 38%, 32% and 28% of the samples, constituting 10.4%, 4.5% and 2.4% of the total fungi, respectively (Table1). This is in agreement with the results obtained from sheep wool in Egypt by Abdel-Gawad (1997) who reported that C. tropicum was the most common species occurring in 58% of the samples. Ali-Shtayeh et al. (1988a) observed that C. keratinophilum and C. tropicum were found respectively in 7.9% and 6.7% of goat hairs from West Bank of Jordan. Deshmukh (2004) reported that C. indicum was the most common species in feathers of pigeon and was represented in 24% of the samples. The above species and other Chrysosporium spp. were also isolated but with different frequencies hairs (Deshmukh. from animal 2004: Mandeell, et al. 2009 and Dokuzeylul et al. 2013).

Aspergillus was the second most frequent genus on camel hair, contaminating 48% of the samples matching 24.8% of total fungi. It was represented by 7 species of which A. flavus and A. niger were the most prevalent species (22 & 32% of samples and 5.7% & 11.7 % of total fungi). A. ustus (12% of the samples), A. terreus (8%), A. ochraceus (6%), A. fumigatus and A. wentii (4% each) were isolated in rare frequency of occurrence (Table 1).In Egypt, Barakat and El-Shanawany (1998) reported that Aspergillus was the second most frequent genus on the hair of donkey. They noticed that A. fumigatus, A. sydowii, and A. versicolor were the most common species. Also, these Aspergillus species have been reported from hairs of camel, cow, donkey and goat in Egypt (Bagy and Abdel-Hafez, 1985; Bagy, 1986), from hair of cows, donkeys, goats, rabbits, cats and dogs in the west bank of Jordan (Ali Shtayeh et al. 1988a,b) and from hair of goats and wool of sheep inYemen (Sallam and Alkolaibe 2010)

In the present study, Scopulariopsis (2 species) occupied the third place in the number of cases of isolation. It was isolated from 28% of samples comprising 8% of total fungi. From the two species isolated S. brevicaulis was the most prevalent while, S. candida was less frequent (Table 1). Several authors reported the prevalence of Scopulariopsis species on hairs of camel (Bagy and Abdel-Hafez, 1985, Nasser et al. 1998; Shokri and Khosravi, 2011) as well as on other animals and birds in Portugal (Bernardo et al. 2005), Bahrin (Mandeell et al. 2009), and Iran (Rostami et al. 2010).

Emericella (2 species), Penicillium (3), Acremonium (2) and Geotrichum (1) were isolated from 7 or 8 samples (out of 50) contributing 3.4%, 2.8%, 3.6% and 1.9% of total fungi, respectively. From the above Ass. Univ. Bull. Environ. Res. Vol. 17 No. 2 October 2014

*genera E. nidulans, P. chrysogenum, A.* strictum and *G. candidum* were the most common species.

Dermatophytes represented by two genera (Microsporum and Trichophyton) were recovered from 7 samples, matching 1.2% and 1.1% of total fungi, respectively. From these genera, M. canis, M. gypseum, T. rubrum and T. verrucosum were identified in rare occurrence. These species were also isolated with different frequencies from hair of different animals as reported by Prado et al. (2008); Madhavi et al. (2011); Gangil et al. (2012); Dokuzeylul et al. (2013) and Mohammed (2013).

Alternaria (2 species), Chaetomium (2), Fusarium (3), Paecillomyces (1) and Thermoascus (1) were isolated in rare frequency of occurrence. They emerged in 8-10% of the samples, accounting collectively 8.6% of total fungi (Table1). These species were also isolated in different frequencies from various keratinous substrates as reported by several authors (Bagy, 1986: Sallam and Alkolaibe, 2010: and Mohammed, 2013).

#### Conclusion

The present study gives an insight on the mycobiota of camel hair in Taiz, Yemen. The prevalence of dermatophytes and nondermatophytes emphasises that camels have a potentiality for shedding fungi in the environment and serve as reservoirs for human pathogens.

Acremonium A. rutilum W. Gams A. strichum W. Gams Alternaria A. alternata (Fr.) Keissler A. tenuissima (Kunze) Wiltshire Aspergillus A. flavus link A. flavus link A. flavus link A. fumigatus Fresenius A. fumigatus Fresenius A. niger van Tieghem A. terreus Thom A. ochraceus Wilhelm A. ochraceus Wilhelm A. ochraceus Wilhelm A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	27 8 19 18 13 5 188 43 5 89 16 14 18 3 24 19 5 311 79 34 173	7 L 3 R 4 R 5 R 3 R 24 M 11 L 2 R 16 M 4 R 3 R 6 R 2 R 5 R 4 R 3 R
A. rutilum W. Gams A. strichum W. Gams Alternaria A. alternata (Fr.) Keissler A. tenuissima (Kunze) Wiltshire Aspergillus A. flavus link A. flavus link A. flavus link A. funigatus Fresenius A. furiegatus Fresenius A. niger van Tieghem A. etrreus Thom A. ochraceus Wilhelm A. ochraceus Wilhelm A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	19 18 13 5 188 43 5 89 16 14 18 3 24 19 5 311 79 34	4 R 5 R 4 R 3 R 24 M 11 L 2 R 16 M 4 R 3 R 6 R 2 R 5 R 4 R 3 R
A. strichum W. Gams Alternaria A. alternata (Fr.) Keissler A. tenuissima (Kunze) Wiltshire Aspergillus A. flavus link A. flavus link A. fumigatus Fresenius A. fumigatus Fresenius A. niger van Tieghem A. terreus Thom A. ochraceus Wilhelm A. ochraceus Wilhelm A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	18 13 5 188 43 5 89 16 14 18 3 24 19 5 311 79 34	5 R 4 R 3 R 24 M 11 L 2 R 16 M 4 R 3 R 6 R 2 R 5 R 4 R 3 R
A. alternata (Fr.) Keissler A. tenuissima (Kunze) Wiltshire Aspergillus A. flavus link A. flavus link A. fumigatus Fresenius A. niger van Tieghem A. terreus Thom A. ochraceus Wilhelm A. ochraceus Wilhelm A. ochraceus Wilhelm A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	13 5 188 43 5 89 16 14 18 3 24 19 5 311 79 34	4 R 3 R 24 M 11 L 2 R 16 M 4 R 3 R 6 R 2 R 5 R 4 R 3 R
A. tenuissima (Kunze) Wiltshire Aspergillus A. flavus link A. fumigatus Fresenius A. niger van Tieghem A. terreus Thom A. ochraceus Wilhelm A. ochraceus Wilhelm A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	5 188 43 5 89 16 14 18 3 24 19 5 311 79 34	3 R 24 M 11 L 2 R 16 M 4 R 3 R 6 R 2 R 5 R 4 R 3 R
Aspergillus A. flavus link A. flavus link A. fumigatus Fresenius A. niger van Tieghem A. terreus Thom A. ochraceus Wilhelm A. ochraceus Wilhelm A. ochraceus Wilhelm A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	188 43 5 89 16 14 18 3 24 19 5 311 79 34	24 M 11 L 2 R 16 M 4 R 3 R 6 R 2 R 5 R 4 R 3 R
A. Javus link A. fumigatus Fresenius A. niger van Tieghem A. terreus Thom A. ochraceus Wilhelm A. ochraceus Wilhelm A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	43 5 89 16 14 18 3 24 19 5 311 79 34	11 L 2 R 16 M 4 R 3 R 6 R 2 R 5 R 4 R 3 R
A. fumigatus Fresenius A. niger van Tieghem A. terreus Thom A. ochraceus Wilhelm A. ustus (Bain.) Thom & Church A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	89 16 14 18 3 24 19 5 311 79 34	16 M 4 R 3 R 6 R 2 R 5 R 4 R 3 R
A. fumigatus Fresenius A. niger van Tieghem A. terreus Thom A. ochraceus Wilhelm A. ustus (Bain.) Thom & Church A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	16 14 18 3 24 19 5 311 79 34	4 R 3 R 6 R 2 R 5 R 4 R 3 R
A. niger van Tieghem A. terreus Thom A. ochraceus Wilhelm A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	14 18 3 24 19 5 311 79 34	3 R 6 R 2 R 5 R 4 R 3 R
A. terreus Thom A. ochraceus Wilhelm A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	18 3 24 19 5 311 79 34	6 R 2 R 5 R 4 R 3 R
A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	3 24 19 5 311 79 34	2 R 5 R 4 R 3 R
A. ustus (Bain.) Thom & Church A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	24 19 5 311 79 34	5 R 4 R 3 R
A. wentii Wehmer Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	19 5 311 79 34	4 R 3 R
Chaetomium C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	5 311 79 34	3 R
C. globosum kunze C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	79 34	A4 **
C. spirale Zopf Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	34	31 H
Chrysosporium C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	-	19 M
C. indicum (Rand. & Sand.) Garg C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	173	16 M
C. keratinophilum D.Frey ex Carm. C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot		23 M
C. tropicum Carmichael C. georgii (Vars. & Ajello) Oorcshot	18	14 M
C. georgii (Vars. & Ajello) Oorcshot	24	6 R
	26 22	8 L 6 R
	4	0 R 2 R
Chrysosporium sp.	13	6 R
Emericella	7	4 R
E. nidulans (Eidam.) Vuill.	3	2 R
E. ruglosa (Thom & Raper) Benjamin	3	2 R
Fusarium	14	7 L
F. verticillioides (Saccardo)Nirenberg	9	7 L
F. oxysporum Schlecht.	6	5 R
F. solani (Mart.) Sacc.	35	2 R 3 R
Geotrichum candidum Link	4	4 R
Microsporum	21	8 L
M. canis Bodin	16	4 R
M. gypsum (Bodin) Gulart & Grigorakis	2	2 R
Mucor hiemalis Wehmer	3	3 R
Paecillomyces lilacinus (Thom) Samson	77	14 L
Penicillium	61	10 L
P. chrysogenum Thom	16 6	6 R 5 R
P. funiculosum Thom	8	5 K 7 L
Penicillium sp.	2	2 R
Scopulariopsis	6	5 R
S. brevicaulis (Sacc.) Bain.	8	7 L
S. candida (Gueguen) Vuillemin		
Thermoascus aurantiacus Miehe		
Trichophyton		
T. rubrum (Castellani) Sabouraud		
T. verrucosum Bodin		
Sterile mycelia		
Gross total counts	759	1
No. of genera	15	+
No. of species	34	

# Table (1): fungi isolated from camel hair (out of 50) samples using soil- plating technique on Saubroud's dextrose agar at 28°C.

TC= Total count; NCI= number of cases of isolation; OR= occurrence remark: H= high occurrence, 25-50 (out of 50) cases; M= moderate occurrence, 13- 24 cases; L= low occurrence, 7- 12 cases; R= rare occurrence, 1-6 cases.

## References

- Abdel-Gawad, K. M. (1997): Mycological and some physiological studies of keratinophilic and other moulds associated with sheep wool. Microbiol. Res. 152: 181-188.
- Ali-Shtayeh, M. S.; Arda, H. M.; Hassouna, M. and Shaheen, S. F. (1988a): *Keratinophilic fungi* on the hair of goats from the West Bank, Jordan. Mycopathologia 104: 103-108.
- Ali-Shtayeh, M. S.; Arda, H. M.; Hassouna, M. and Shaheen, S. F. (1988b): *Keratinophilic fungi* on the hair of cows, donkey, rabbits, cats and dogs from the West Bank Jordan. Mycopathologia 149: 193-121.
- Ali-Shtayeh, M. S.; Arda, H. M.; Hassouna, M.; and Shaheen, S. F. (1989): *Keratinophilic fungi* on sheep hairs from West Bank of Jordan. Mycopathologia 106: 95-101
- Bagy, M. M. K. and Abdel- Hafez, A. I. I. (1985): Mycoflora of camel and goat hairs from Al- Arish, Egypt. Mycopathologia 92: 125-128.
- Bagy, M .M .K. (1986): Fungi on the hair of large mammals in Egypt. Mycopathologia 93: 73-75.
- Barakat, A. and El-Shanawany, A. A. (1998): Studies on the mycoflora associated with donkey hair at Assiut. Assiut Vet. Med. J. 8(15-17): 283-294.
- Bernardo, F.; Lanca, A.; Guerra, M. M. and Martins, H. M. (2005): Dematophytes

- isolated from pet, dogs and cats, in Lisbon, Portugal (2000-2004). Rev. Port. Cienc. Vet. 100: 85-88.
- De Hoog G. S., Guarro J., Gene J. and Figueras M. J. (2000): Atlas of Clinical Fungi. Centraalbureau voor Schimmellcultures, Universitat Rovira I Virgili. Reus. Spain.
- Deshmuka, S. K. (2004): *Keratinophilic fungi* on feathers of pigeon in Maharashtra, India. Mycoses 47: 213-215.
- Dobrowolskh, A.; Staczek, P.; Kaszuba, A. and Kozlowska, M. (2006): PCR-RFLP analysis of the *dermatophytes* isolated from patient in central Poland. J. Dermatological Science 42: 71-74.
- Dokuzeylul, B.; Kahraman, B. B. and Sigirci,
  B. D. (2013): *Dermatophytosis* caused by a *Chrysosporium species* in two cats in Turkey: a case report. Veterinarni Medicina 58: 633-636.
- El-Said, A. H. M.; Sohair, T. H. and El-Hadi, A. G. (2009): Fungi associated with the hairs of goat and sheep in Libya. Mycobiology 37(2): 82-89
- Gangil, R.; Dutta, P. and Tripathi, R. (2012): Incidence of *dermatophytosis* in canine cases presented at Apollo Veterinary College, Rajashtan, India. Vet. World 5(11): 682-684.
- Khanam, S. J. P. and Jain, P. C. (2002): Isolation of keratin degrading fungi from soil of Damoh, India. Asian J. Microbiol. Biotechnol. Environ. Sci. 4: 251-254.

- Kraemer, A.; Mueller, R. S.; Werckenthin, C.; Straubinger, R. K. and Hein, J. (2012): *Dermatophytes* in pet Guinea pigs and rabbits . Veterinary Microbiology 157: 208-213.
- Madhavi, S.; Rama, R. M. V. and Jyothsna, K. (2011): Mycological study of *dermatophytosis* in rural population. Annuals Biological Research 2(3): 88-93.
- Maghraby, A. T.; Gherbawy, Y. A. M. H. and Hussein, A. M. (2008): *Keratinophilic fungi* inhabiting floor dusts of student houses at South Valley University in Egypt. Aerobiologia 24: 99-106.
- Mandeel, Q.; Nardoni, S. and Mancianti, F. (2009): *Keratinophilic fungi* of feathers on common clinically healthy birds in Bahrain. Mycoses 54: 71-77
- Marcella, R.; Mercantini, R.; Spinelli, P. and Volterra, L. (1985): Occurrence of *keratinophilic fungi* in animals of the zoological park of Roma. Mykosen 28: 507-512.
- Marcella, R. and Mercantini, R. (1986): *Keratinophilic fungi* isolated from soils of the Abruzzo National Park, Italy. Mycopathologia 94: 97-107.
- Mohammed, S. J. (2013): Dermatophytes isolated from dogs suspected of dermatophytosis in Baghdad City. Diyala J. Pure Sci. 9(4): 61-66.
- Moses, O. E. and Sunday, O. F. (2001): Occurrence of *keratinophilic fungi* and dermatophytes on birds in Nigeria. Mycopathologia 153: 87-89.

- Moss, E. S. and McQuown, A. I. (1969): Atlas of medical mycology 3rded. The Williams and Wilkins Company, Baltimore U.S.A.
- Moubasher A. H. (1993): Soil fungi in Qatar and other Arab countries. Scientific and Applied Research Center. University of Qatar,566 pp.
- Mukesh, S. and Sharma, M. (2010): Incidence of dermatophytes and other keratiophilic fungi in schools and college playground soils of Jaipur, India. Afr. J. Microbial. Research 4(24): 2647-2654.
- Nasser, L. A.; El-Shanawany, A. A. and Barakat, A. (1998): Ecological and physiological studies on fungi associated with camel hairs from Saudi-Arabia. Assiut Vet. Med. J. 8(15-17): 243-254.
- Nichita, I. and Marcu, A. (2010): The fungal microbiota isolated from cats and dogs. Animal Science and Biotechnologies 43(1): 411-414.
- Ogbonna, C. I. C.; Enweani, I. B. and Ogueri, S. C. (1986): The distribution of ringworm infections amongst Nigerian nomadic fulani herdsmen. Mycopathologia 96: 45-51.
- Periasamy, A.; Hilda, A. and Subash, C. B. G. (2004): *Keratinophilic fungi* of poultry farm and feather damping soil in Tamil Nadu, India. Mycopathologia 158: 303-309.
- Prado, M .R.; Brilhante, R .S. M. and Cordeiro, R. A. (2008): Frequency of yeasts and dermatophytes from healthy

Ass. Univ. Bull. Environ. Res. Vol. 17 No. 2 October 2014

and diseased dogs. J. Vet. Diagn. Invest. 20: 197-202.

- Ress, R. G. (1967): *Keratinophilic fungi* from Queensland. 11. Isolation from animal hairs and scales. Sabouraudia 5: 165-172.
- Rippon, J. W. (1982): The pathogenic fungi and the pathogenic *actinomycetes*. Philadelphia: W. B. Saunders Company.
- Rostami, A.; Shirani, D.; Shokri, H.; Khosravi,
  A. R.; Daieghazvini, R. and Tootian, Z.
  (2010): Fungal flora of the hair coat of Persian squirrel (Sciurus anomalus) with and without skin lesion in Tehran, Iran.
  J. de Mycologie Medicale 20: 21-25.
- Sallam, A. M. H. and Alkolaibe, A. M. (2010): Distribution pattern of other *keratinophilic fungi* on goats hair and sheep wool,Taiz city, Yemen. J. Environmental Sciences 39 (3): 345-356.
- Shokri, H. and Khosravi, A. R. (2011): Fungal

flora isolated from the skin of healthy dromedary camels Camelus dromedaries. Int. J. Vet. Res. 5 (2): 109-112.

- Shukia, P.; Shukla, C. B.; Kango, N. and Skukla, A. H. (2003): Isolation and characterization of a dermatophyte, Microsporium gypseum from soils of Rewa (Madhya Pradesh), India. Pak. J. Bio. Sci. 6: 622-625.
- Singh, I.; Mishra, A. and Kushwaha, R. (2009): Dermatophytes, related keratinophilic and opportunistic fungi in indoor dust of houses and hospitals. Indian J. Med. Microbial. 27: 242-246.
- Ulfig, K. (2007): Influence of peptone, ammonia, water and urea supplements on *keratinolytic* and associated non*keratinolytic fungi* in sewage sludge. Int. Biodeterioration & Biodegradation 59: 62-68.

Ass. Univ. Bull. Environ. Res. Vol. 17 No. 2 October 2014

# الملخص العربى

# الفطريات المصاحبة لشعر الجمال بمدينة تعز- اليمن

على محمد حسن سلام \* جمال العامري \* \*

قسم الميكروبيولوجى التطبيقى - كلية العلوم \* قسم الميكروبيولوجي كلية الطب \* \* - جامعة تعز - اليمن

استهدف البحث التعرف على الأحياء الفطرية لخمسين عينة من شعر الجمال جمعت من أماكن مختلفة من مدينة تعز وذلك باستخدام طريقة مصائد التربة والوسط الغذائي سبارود اجار والتحضين عند درجة حرارة ٢٨°م وقد تم عزل ٣٤ نوعاً فطرياً تنتمي إلى ١٥ جنساً وكان أكثر الأجناس شيوعاً وانتشاراً هو جنس كريزوسبورم يليه اسبرجلس ثم جنس اسكوييولاريوييسس كما تم أيضاً عزل أنواع من الفطريات الجلدية تنتمي إلي الميكروسبورم والتريكوفيتون ولوحظ أن العديد من الفطريات المعزولة يمكن أن تكون ممرضة للإنسان.